Biomedical Applications of Polycaprolactone (PCL) Composites: Structure, Properties, and Future Prospects

Prajakta Subhedar*, Divya Padmanabhan and Richa Agrawal

Department of Mechanical Engineering, Pillai College of Engineering, Navi Mumbai-410206, India

Abstract: Polycaprolactone (PCL) is a semi-crystalline, biodegradable aliphatic polyester that has emerged as a versatile biomaterial for tissue engineering, drug delivery, and regenerative medicine applications due to its exceptional biocompatibility, controlled degradation kinetics (2-4 years in vivo), and FDA approval status for multiple medical devices. Despite these advantages, pure PCL exhibits significant limitations including low mechanical strength (16-24 MPa tensile strength), hydrophobic surface properties (water contact angle 80-90°), and minimal bioactivity, which restrict its clinical utility in load-bearing and cell-interactive applications. To address these shortcomings, researchers have developed PCL-based composite systems by incorporating bioactive ceramics (hydroxyapatite, β -tricalcium phosphate), natural polymers (collagen, chitosan, gelatin), synthetic polymers (PLA, PLGA), and nanomaterials (carbon nanotubes, graphene oxide) to create multifunctional biomaterials with enhanced properties. This comprehensive review analyzes PCL composite development over the past two decades, emphasizing fabrication techniques including electrospinning, 3D printing, solvent casting, and melt blending, which enable precise control over scaffold architecture and functionality. Comparative analysis with other biodegradable polymers (PGA, PLGA) reveals PCL's unique advantages in long-term applications, with studies demonstrating >90% cell viability, ~65% bone regeneration in animal models, and sustained drug release profiles extending 6-8 weeks. Recent innovations include smart, stimuli-responsive PCL systems for targeted therapy, gene delivery platforms, and bioprinting applications that have advanced from laboratory research to clinical trials, with several PCL-based products (Neurolac®, Osteoplug®) receiving regulatory approval. Current challenges include manufacturing scalability, long-term biocompatibility assessment, and complex regulatory pathways for multi-component systems. Future developments focus on integrating artificial intelligence for scaffold design, 4D printing technologies for dynamic structures, and multidisciplinary approaches combining materials science with precision medicine. This review demonstrates that PCL-based composites represent a transformative class of biomaterials with customizable properties that bridge fundamental research and clinical translation, positioning them at the forefront of next-generation biomedical technologies.

Keywords: Biodegradable composites; biomedical applications; drug delivery systems; Polycaprolactone, tissue engineering.

1. INTRODUCTION

In recent years, biodegradable polymers have become indispensable in biomedical research and clinical practice. Their ability to break down into harmless metabolites not only eliminates the need for secondary removal surgeries but also minimizes chronic inflammatory responses [1,2]. These advantages have led to their widespread adoption in drug delivery, regenerative medicine, and tissue engineering [3].

Among the different candidates, polycaprolactone (PCL) has attracted strong interest due to its rare combination of biocompatibility, flexibility, and a very slow degradation rate [4]. These features make it particularly useful in applications where prolonged stability is required, such as bone fixation devices, long-term scaffolds, and controlled drug release carriers. Additionally, PCL is highly adaptable to different fabrication techniques, including electrospinning and additive manufacturing, which enable the creation of complex architectures suited for patient-specific therapies [5].

Despite these benefits, PCL on its own presents shortcomings that limit its broader clinical utility. Its

*Address correspondence to this author at the Department of Mechanical Engineering, Pillai College of Engineering, Navi Mumbai-410206, India;

E-mail: prajakatasubhedar@gmail.com

hydrophobic nature hinders early-stage cell attachment, while its relatively low strength restricts application in load-bearing conditions. Furthermore, being biologically inert, it provides minimal biochemical cues for tissue regeneration. To address these barriers, considerable research has focused composite systems where PCL is blended or reinforced with bioactive ceramics, natural polymers, nanomaterials. Such modifications degradation behavior, improve surface interactions, and increase mechanical reliability [6,7]. Recent reports also highlight how advanced fabrication technologies have expanded the scope of PCL-based composites, offering more precise control over architecture and functionality [8,9].

2. POLYCAPROLACTONE (PCL): PROPERTIES AND BIOMEDICAL RELEVANCE

semicrystalline PCL is aliphatic polyester synthesized via ring-opening polymerization of εcaprolactone using catalysts like stannous octoate. Its linear structure contributes to its crystallinity and mechanical properties [10,11]. Polycaprolactone (PCL) exhibits a distinct set of physicochemical properties, including controlled biodegradability, mechanical strength, thermal stability, and surface wettability, which collectively define its performance in biomedical and environmental applications.

E-ISSN: 1929-5995/25

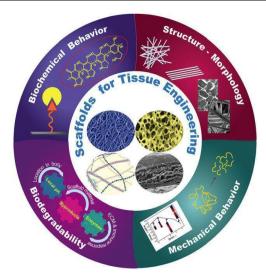


Figure 1: Elements of biomaterial scaffolds that are essential for inducing the behavior of cells.

2.1. Physicochemical Properties

2.1.1. Biodegradability

PCL degrades slowly through hydrolysis of its ester linkages, suitable for long-term applications. It exhibits excellent biodegradability, making it highly suitable for biomedical and environmental applications. The rate of degradation can be tailored by modifying molecular weight or blending PCL with other polymers [4,12]. Due to its controlled degradation profile, PCL is widely used in long-term drug delivery systems, tissue engineering scaffolds, and bioresorbable implants [6].

2.1.2. Mechanical Strength

PCL possesses moderate mechanical strength, which makes it suitable for soft and flexible biomedical applications. It has good elongation at break and ductility, allowing it to deform without cracking under stress [13]. However, its low tensile and compressive strength limit its use in load-bearing applications without reinforcement. The mechanical performance of PCL can be adjusted by controlling its molecular weight and crystallinity [14]. Despite its limitations, PCL's mechanical properties are adequate for applications like soft tissue engineering, wound dressings, and controlled drug delivery systems [15].

2.1.3. Thermal Stability

PCL exhibits a low melting point (~60°C), allowing easy processing. It also demonstrates good thermal stability, with a melting temperature typically around 58-63 °C [16]. It maintains its structural integrity during common polymer processing methods such as extrusion, electrospinning, and 3D printing. PCL begins to thermally degrade at temperatures above 200 °C, making it suitable for applications requiring moderate heat resistance [17]. This thermal behavior supports its versatility in biomedical device fabrication and drug delivery systems [18].

2.1.4. Wettability and Water Contact Angle

PCL exhibits relatively low wettability due to its hydrophobic surface, typically showing a water contact angle between 80° and 90° [19,20]. This moderate to high contact angle can limit initial cell adhesion and protein adsorption on its surface. To enhance its biomedical performance, surface treatments such as plasma modification or blending with hydrophilic polymers are often applied to reduce the contact angle. Improved wettability of PCL supports better cell attachment, proliferation, and overall biocompatibility in tissue engineering applications [21,22].

2.1.5. Comparative Analysis with Other Biodegradable Polymers

To better understand PCL's position among biodegradable polymers used in biomedical applications, a comparative analysis with polyglycolic acid (PGA) and poly(lactic-co-glycolic acid) (PLGA) is essential. These three polymers represent the most widely utilized biodegradable materials in tissue engineering and drug delivery systems, each offering distinct advantages based on their molecular structure and degradation characteristics.

This comparative analysis demonstrates that while PGA offers rapid degradation and high mechanical strength, and PLGA provides tunable properties through copolymer ratios, PCL's extended degradation timeline and processing flexibility make it particularly suitable for long-term biomedical applications requiring sustained performance over months to years.

2.2. Biological Properties

2.2.1. Biocompatibility and Biostability

PCL exhibits excellent cytocompatibility, with fibroblast and osteoblast cultures showing >90% cell viability after 7 days and enhanced proliferation on

Property	PCL	PLA	PGA	PLGA
Molecular weight [g/mol]	3,000-80,000 [10,11]	10,000-300,000 [23,28]	20,000-100,000 [23,24]	10,000-200,000 [25,26]
Density [g/cm³]	1.07-1.15 [16]	1.21-1.25 [23,28]	1.50-1.70 [23,24]	1.20-1.30 [25,26]
Tensile strength [MPa]	16-24 [4, 13]	50-70 [23,28]	60-99 [23,24]	40-55 (composition- dependent) [25,26]
Young's modulus [GPa]	0.21-0.44 [4,13,14]	2-3 [23,28]	6-7 [23,24]	1-2 [25,26]
Crystallinity [%]	~69 [4,16]	37-40 [23,28]	~45-55 [23,24]	Amorphous to semi- crystalline (varies with ratio) [25,26]
Glass transition temperature [°C]	-65to -60 [4,17]	55-65 [23,28]	35-40 [23,24]	45-55 [25,26]
Melting temperature [°C]	56-65 [16,17]	170-180 [23,28]	225-230 [23,24]	No sharp melting point [25,26]
Decomposition temperature [°C]	~350 [16,17]	~250-280 [23,28]	~260-280 [23,24]	~200-250 [25,26]

Table 1: Comparative Properties of Biodegradable Polymers: PCL, PGA, and PLGA

electrospun scaffolds [29,30]. *In vivo*, PCL–HA composites achieved ~65% bone regeneration in rat calvarial defects within 8 weeks [31]. Subcutaneous implantation studies reported structural stability for over 12 months with only mild local responses, while degradation products (6-hydroxycaproic acid) were safely metabolized via the citric acid cycle [32]. These outcomes confirm PCL's suitability for long-term biomedical applications such as bone regeneration, vascular grafts, and extended drug delivery.

2.2.4. FDA Approval Status

PCL holds FDA approval for use in various medical applications, including drug delivery devices, absorbable sutures, adhesion barriers, and aesthetic fillers like microsphere-based collagen stimulators. Its regulatory clearance spans both implantable and resorbable medical devices, underscoring its safety profile and compatibility with human tissues [33-36]. The FDA has incorporated PCL into its Material Safety Summaries, reflecting its long-term stability and biocompatibility when used in clinical settings. Publicly available safety assessments note minimal systemic reactions and primarily mild, device-specific, local responses-such as transient inflammation or seroma formation-aligned with other approved biomaterials [23,37].

3. LIMITATIONS OF PURE PCL IN BIOMEDICAL APPLICATIONS

Despite its widespread use in biomedical applications, pure polycaprolactone (PCL) possesses several limitations that restrict its effectiveness in certain clinical settings, given below.

3.1. Slow Degradation Rate

Pure PCL exhibits a significantly slow degradation rate under physiological conditions, often taking

several months to years to fully resorb in the body. While this can be beneficial for long-term applications, it poses limitations in cases where rapid tissue regeneration or drug release is required. The prolonged presence of the material may lead to chronic inflammation or impede natural tissue remodeling in fast-healing tissues such as skin or muscle [38,39].

3.2. Hydrophobicity

PCL has a relatively high water contact angle, indicating its hydrophobic nature. This poor wettability limits its ability to support initial protein adsorption and cell adhesion, both of which are critical for effective tissue integration. The hydrophobic surface can hinder cellular proliferation and delay healing processes, especially in applications like wound healing or tissue scaffolds [40,41].

3.3. Mechanical Constraints

While PCL is flexible and ductile, it lacks sufficient mechanical strength for load-bearing applications such as bone or joint implants. Its low tensile and compressive strength restrict its use in high-stress environments without reinforcement. Additionally, pure PCL can deform under sustained mechanical load (creep), which compromises its long-term structural reliability in orthopedic and dental applications [4,42].

3.4. Limited Bioactivity

Pure PCL is biologically inert, meaning it does not actively interact with surrounding tissues to stimulate healing or regeneration. It lacks inherent cues for osteoconduction or angiogenesis, which are essential for applications such as bone regeneration or vascular grafts. This limited bioactivity necessitates the addition of bioactive fillers (e.g., hydroxyapatite, growth factors) to enhance its functionality [43,44].

3.5. Limited Antibacterial Properties

Polycaprolactone (PCL) by itself does not possess inherent antibacterial properties, which may limit its effectiveness in infection-prone biomedical applications. However, its polymer matrix can be easily modified to incorporate antibacterial agents such as silver nanoparticles, antibiotics, or natural extracts. These modifications help inhibit bacterial growth on implant surfaces, reducing the risk of post-surgical infections [45].

These limitations highlight the need for modifying or reinforcing PCL through blending, surface functionalization, or composite formation to expand its utility across a broader range of biomedical applications.

4. PCL-BASED COMPOSITES: NEED AND STRATEGIES

4.1. Why Blend or Reinforce PCL?

Blending or reinforcing PCL addresses its shortcomings by enhancing bioactivity, mechanical strength, and degradation behavior. Despite the benefits of PCL, its slow degradation rate, hydrophobicity and limited bioactivity remain significant barriers in clinical use [23,46]. Blending or copolymerizing PCL with other constituents alters the scaffold's functional properties. In this regard, different studies have combined distinct polymers, metals, and ceramics with PCL to increase its properties [47].

The glass transition temperature (Tg) of PCL is 213 K, while its melting temperature (Tm) is between 332 and 337 K [48]. It blends well with other polymers and has a low melting point, making it ideal for scaffolding and drug delivery systems. Although PCL has sufficient biocompatibility on its own, enhanced bioactivity is preferred for a few uses [49]. PCL can be made more bioactive by either copolymerization or the addition of bioactive fillers. The co-polymerization, kind of biopolymer link, monomer type, and adjacent groups all influence hydrolysis and consequently the rate of breakdown [55]. Copolymerization and blending have both been utilized to slow the degradation of polymeric scaffolds. According to research, integrating polymers and/or ceramics with PCL may help alter the rate of deterioration while keeping the functional qualities of the scaffold [50].

PCL has a hydrophobic tendency in general. Hydrophilicity and improved cellular compatibility are required for it to function as a scaffold. Multiple efforts have been made to alter the exterior PCL constructs by covering them with gelatin, proteoglycans, growth factors, fibrin, and fibronectin. PCL scaffolds with

double protein coatings promote proliferation, colonization, and cell adhesion. However, because of the surface's ECM matrix rearrangement, the arginine-glycine-aspartic acid (RGD) coating showed smoother surfaces and early bone deposition onto scaffold surfaces [51,52].

4.2. Types of Reinforcements Used

Natural fibers such as collagen, chitosan, and cellulose, ceramic fillers like hydroxyapatite (HA) [53,54] and β -tricalcium phosphate (β -TCP) [55,56] are commonly used as reinforcements in PCL composites to enhance biocompatibility and mimic the extracellular matrix. Carbon-based materials, including graphene oxide [57] and carbon nanotubes [58], are used to enhance the mechanical strength and electrical conductivity of PCL composites. Synthetic polymers such as polylactic acid (PLA) [59] and polyglycolic acid (PGA) [23] are blended with PCL to tailor degradation rates and mechanical performance. Synthetic polymers like poly-lactic co-glycolic acid (PLGA) [60], polyglycolic acid (PGA), poly-L-lactic-acid (PLLA) [61], polylactic acid (PLA) [62], graphene [63], and gelatin (Ge) [64] are used to make blended composites, and ceramics such as hydroxyapatite (HA), \(\beta\)-tricalcium phosphate (β-TCP), zinc oxide (ZnO), titanium oxide (TiO₂), bioglassetc [65,66].

5. PROCESSING TECHNIQUES FOR PCL COMPOSITES

To make blended PCL composites, several technologies such as electrospinning, melt blending and extrusion, solvent casting, 3D printing, and melt electrospinning can be employed [67,68]. fabrication method is determined by the type of scaffold, such as porous or fibrous. These techniques influence the final morphology, porosity, mechanical strength, and degradation behavior of the composites. which are crucial for biomedical applications. Solvent casting is often used for simple film formation, whereas melt blending is favored for thermoplastic formulations. Electrospinning and 3D printing provide high-resolution tailored tissue scaffolds for engineering regenerative medicine [69].

5.1. Electrospinning

Electrospinning is a widely used technique to fabricate nanofibrous scaffolds from PCL composites for biomedical applications. It enables the production of fibers with high surface area and porosity, closely mimicking the structure of natural extracellular matrix [70,71]. By incorporating bioactive materials or other polymers into the PCL solution, electrospun composites can achieve enhanced mechanical strength and biological functionality. This method allows for controlled fiber diameter and alignment, which are important for guiding cell behavior and tissue regeneration. As a result, electrospun PCL composites are extensively used in wound healing, tissue engineering, and drug delivery systems [72].

5.2. Solvent Casting

Solvent casting is a simple and effective method used to fabricate PCL composites into thin films or membranes for biomedical applications. In this technique, PCL and reinforcing agents or bioactive fillers are dissolved in a suitable solvent and cast onto a flat surface, followed by solvent evaporation. This process allows uniform dispersion of additives, improving the composite's mechanical, thermal, and biological properties [64,73]. Solvent casting is ideal for incorporating drugs, nanoparticles, or natural polymers to enhance biocompatibility and functionality. PCL composite films produced by this method are commonly used in wound dressings, drug delivery systems, and barrier membranes [74].

5.3. Melt blending and extrusion

Melt blending and extrusion are widely used thermal processing techniques for fabricating PCL composites with improved mechanical and functional properties. In this method, PCL and reinforcing materials such as ceramics, polymers, or nanoparticles are mixed at elevated temperatures without using solvents, making the process environmentally friendly [75,76]. The molten mixture is then extruded into desired shapes like filaments, rods, or sheets suitable for biomedical applications. This technique ensures good dispersion of fillers and allows for scalable production with controlled composition and geometry. Melt-blended PCL composites are commonly used in orthopedic implants, scaffolds, and 3D printing filaments [76].

Table **2** summarizes the effect of different compositions and fabrication techniques on the functional performance of PCL-based scaffolds, highlighting how material combinations and processing routes influence their mechanical properties, degradation behavior, and biomedical applications.

5.4. 3D Printing/Additive Manufacturing

3D printing, or additive manufacturing, is a versatile technique used to fabricate complex and customized structures from PCL composites for biomedical applications. It allows precise control over scaffold architecture, porosity, and geometry, which is crucial for tissue engineering. By incorporating bioactive fillers or reinforcing agents into PCL, the printed composites

can exhibit enhanced mechanical strength and biological performance.

This method supports the creation of patient-specific implants and drug delivery devices with tailored degradation profiles. PCL composites processed through 3D printing are widely applied in bone regeneration, cartilage repair, and soft tissue engineering. SLA, SLS, FDM, and bioprinting are some of the additive manufacturing (AM) processes utilized to create PCL-based scaffolds. These methods are reviewed in Table 3.

6. BIOMEDICAL APPLICATIONS OF PCL-BASED COMPOSITES

Polycaprolactone-based composites have emerged as versatile platforms for various biomedical applications due to their biodegradability, tunable degradation, and ability to be processed into functional scaffolds [4, 28]. The key application areas are outlined below.

6.1. Tissue Engineering

PCL-based scaffolds have shown promising outcomes in bone, cartilage, and skin regeneration due to their tunable mechanics and biodegradability [15,31,53,54]. In a rabbit calvarial defect model, PCL/HA composites achieved ~65% new bone formation within 8 weeks, significantly outperforming pure PCL (~35%) [31,53]. Electrospun PCL nanofibers seeded with chondrocytes demonstrated a twofold increase in glycosaminoglycan (GAG) production compared to PLA-based scaffolds, indicating superior cartilage matrix deposition [30,59,62]. For skin repair, porous PCL dressings accelerated wound closure by ~40% compared to untreated controls, with improved angiogenesis and collagen deposition [51,64,74]. Vascular graft studies further report endothelialization rates above 80% within 30 days, comparable to or better than PLGA conduits [61,77].

6.2. Drug Delivery Systems

PCL carriers enable sustained drug release through a biphasic profile: an initial burst (surface drug) followed by diffusion- and degradation-controlled release lasting weeks to months [6,32,46]. For PCL nanoparticles instance, doxorubicin-loaded released ~30% in 24 h and ~85% over 30 days, while gentamicin-loaded microspheres maintained activity for >6 weeks [6,45,60]. Hydrophobic drugs (e.g., paclitaxel) show high encapsulation (>80%) and slower release due to strong affinity with PCL [4,49], whereas hydrophilic agents require stabilizers to avoid rapid release [46,50]. These features make PCL particularly effective for long-term cancer therapy, wound care, and growth factor delivery [6,44,45].

Table 2: Effect of composition and fabrication Methods on Functional Characteristics of PCL Scaffold

Composite & configuration	Fabrication process used	Effect on functional properties	Degradation	Application	Ref.
PCL/rhBMP-2/HA	Extrusion based 3D printing	Enhanced the angiogenesis, osteoblastic differentiation of stem cells, and new bone formation.	-	Dental reconstructive surgery	[31,53,54]
PCL/PLGA 0.1:0.9,0.5:0.5, 0.9:0.1	3D printing	High porosity in 50:50 scaffold with high mechanical properties supporting load bearing application.		Scaffold for human periodontal ligament stem cells application	[46,60]
PCL/PGA/HA PGA (0 & 25%) HA (0,10,20%)	Solvent extraction, extrusion based 3D printing	Enhanced mechanical properties, increased hydrophilicity & cell adhesion.	Enhanced degradation	Bone tissue engineering and resorbable barrier membrane	[4,42,55]
PCL/Ge/nHA ((70:30):15)	Electrospinning	Increased ALP activity, proliferation, cell adhesion, and mineral deposit on scaffold.	Faster degradation of scaffold	Bone tissue engineering	[51,52,64]
PCL/CA & PCL/CA/LA	Electrospinning	Enhanced cell spreading and proliferation	-	Regenerative medicine and tissue engineering applications	[47,64]
PCL	Solvent casting & fused deposition modeling	Increased stiffness, crystallinity	1%-7% degradation & degradation is non-toxic at molecular level	Long term (2yrs) use and bone generation	[38,39,74]
PCL/SrHA 0,10,15%	Electrospinning		Degradation rate: PCL < PCL/SrHA 10 % wt< PCL/SrHA 15 % wt.	Tissue engineering	[53,54,77]
PCL/PLA 16:4,18:2,9:1	Two photon polymerization	Higher content of PCL results in higher compressive modulus but gradual decrease in compressive strength as a result of progressive mass loss over time.	Degradation rate: 16:4<18:2<9:1 Slower degradation as compared to PLA	Regenerative medicine and tissue engineering applications	[59,62]
PCL/PLLA & PCL/PCLA 50:50	Dual cylinder chamber molding		PCL/PLLA degradation rate is intermediate and increases with PLLA content	Tissue-engineered vascular grafts	[61,77]
PCL/β TCP (E beam treated) 100:0,80:20,60:40, 40:60	Dissolution and precipitation phases, Pre-E-Beam Surface Treatment	-	Accelerated degradation due to E beam treating	Subcutaneous implantation	[55,56,58]
PCL/β-TCP 80:20	Extrusion based 3D printing	Controlled porosity, improved mechanical properties	Increased degradation rate 60% porous material showed mass loss of 40% at 48 hrs	Osteonecrosis of the femoral head	[53,55,76]
PCL/Gelatin 70:30	Electrospinning	-	Degradation rate: PCL <pcl gelatin<="" td=""><td>Skin tissue engineering application</td><td>[51,64,74]</td></pcl>	Skin tissue engineering application	[51,64,74]
PCL/HA/ZnO (ZnO 1,3,6%)	Electrospinning	Increased ALP activity followed by mineralization	ZnO accelerates scaffold degradation	Mid- and long-term resorption for accelerated bone tissue regeneration	[65,66,78]
PCI/TiO2	Microsphere sintering	Good cell differentiation leading to ECM mineralization		Load-bearing bone tissue engineering applications	[63,77,79]

Table 2 (Continue)

Composite & configuration	Fabrication process used	Effect on functional properties	Degradation	Application	Ref.
PCL coated TCP/Bioglass	lost sponge foam method	Formation of apatite on scaffold's surface	Coated specimen showed higher degradation than the uncoated scaffolds	Bone/tissue engineering	[65,66,80]
PCL/PTMC (1:1,1:3,3:1)	Electrospinning	Increased macrophage- mediated foreign body reaction	Degradation speed increases along with PTMC	-	[50,81,82]

Table 3: 3D Printing Methods for PCL Based Composites

3D printing process	Advantages	Limitations	Ref.
SLA	High resolution, high precision, manufacturing of complicated shapes	Difficult to produce a micron-sized scaffold,photo initiator-induced cytotoxicity, limited layer thickness, expensive	[83,84]
SLS	Controlled porosity and pore size	Non-load bearing scaffolds, expensive, involves high temperatures	[85,86]
FDM	Easy to use, economical, controlled functional properties	Restricted resolution, involves high temperatures	[44,69,87]
Binder jet printing	Compatible with a range of materials, economical & fast	Limited resolution and direction of injection, low cell density	[88-90]

6.3. Wound Healing

Electrospun PCL dressings loaded with silver nanoparticles or antibiotics improve antibacterial protection and maintain moisture balance at the wound site [5,45,64]. Growth-factor-incorporated scaffolds enhance angiogenesis and accelerate tissue repair [53,72,74].

6.4. Dental and Orthopedic Applications

Guided bone regeneration membranes made from PCL prevent soft tissue ingrowth and support bone repair [31,53,54]. PCL reinforced with HA or β -TCP mimics natural bone mineral and improves osteointegration in dental implants [55,56,76].

6.5. Nerve Regeneration and Neural Interfaces

PCL conduits provide structural guidance for axonal regrowth across nerve gaps [50,61]. While incorporation of conductive materials (e.g., CNTs, graphene) enhances electrical stimulation for neural repair [58,59,63]. These composites are also being explored in bioelectronic neural interfaces for signal transmission [57,58,77].

6.6. Environmental and Packaging Applications

Beyond biomedicine, PCL has gained attention in sustainable packaging and environmental remediation [18,22,23]. Under industrial composting conditions, PCL films typically degrade within 6-12 months, while

in soil or aquatic environments, complete mineralization may require 2-3 years, depending on crystallinity and thickness [4,16,38]. In comparison, PLA shows faster degradation in compost (~3-6 months) but slower breakdown in soil due to lower microbial activity [23,28,62].

Lifecycle assessment (LCA) studies further highlight PCL's advantages: production generates up to 30-40% lower CO₂ emissions than conventional polyethylene when bio-based raw materials are used [27,73]. Moreover, PCL blends with starch or PLA not only accelerate degradation but also reduce environmental footprint without compromising mechanical properties [59,62,64].

7. RECENT ADVANCES AND INNOVATIONS

Recent innovations in PCL-based composites focus on creating multifunctional materials for advanced biomedical technologies. Smart composites are engineered to respond to physiological cues like pH and temperature, enabling controlled and site-specific drug delivery [91,92]. For example, PCL blended with thermo-responsive polymers can trigger drug release in response to temperature changes, while pH-sensitive versions target acidic tumor environments [99]. Another key area is targeted therapy, where PCL surfaces are functionalized with biomolecules to deliver therapies directly to specific cells [92,93]. PCL scaffolds are also a frontier in gene delivery, providing localized and sustained release platforms for genetic

Figure 2: Biomedical applications of PCL composites.

materials like DNA, siRNA, and CRISPR/Cas9 components to promote tissue regeneration and treat diseases [93,94].

7.1. Al-Driven Scaffold Design and Machine Learning Applications

The integration of artificial intelligence (AI) and machine learning (ML) has become a transformative approach in PCL composite design. Predictive models complex algorithms to optimize architecture and processing parameters, significantly reducing development time and costs [95]. Al-guided methods can predict a scaffold's mechanical properties and refine 3D printing parameters for improved uniformity [95,96]. Additionally, ML models can forecast a composite's performance, degradation and drug release kinetics, before physical testing [96]. By analyzing vast datasets and scientific literature, these computational tools are accelerating knowledge discovery, making AI an indispensable part of next-generation PCL development [95,96].

7.2. Clinical Translation and Market Impact

In terms of clinical translation, PCL-based products have successfully bridged the gap from laboratory research commercial application. to Several bioresorbable medical devices, such as the Neurolac® nerve conduit and the Osteoplug® cranial implant, have gained regulatory approvals after demonstrating their safety and efficacy in clinical trials, including a randomized controlled trial (NCT01119144) that found a PCL/Tricalcium Phosphate implant to be a safe alternative to titanium [97]. This clinical success is a major catalyst for the PCL market, which is projected to grow from its 2022 valuation of approximately \$950 million to over \$1.1 billion by 2030 [97]. Driven by a robust compound annual growth rate (CAGR) of nearly

10%, the healthcare sector is a primary force in this expansion, confirming the strong market penetration and commercial viability of these advanced bioresorbable devices [97].

PCL's cost-effectiveness is a key factor in its commercial success. Compared to other biomedical polymers and traditional materials, its relatively low production cost makes it an attractive option. This economic advantage, combined with its favorable properties and the potential to reduce long-term healthcare costs by avoiding implant removal surgeries, positions PCL as a commercially viable and competitive material in the growing medical device market.

8. CHALLENGES AND FUTURE PERSPECTIVES

Despite the growing promise of PCL-based composites in biomedical applications, several key challenges must be addressed to enable their broader clinical translation and commercial success.

8.1. Scalability and Reproducibility

While laboratory-scale fabrication techniques allow for precise control over composition and structure, scaling up these processes often results in variability in material properties, porosity, and drug loading efficiency [98,99]. This inconsistency hampers large-scale production and limits the feasibility of standardized medical devices and implants. Advanced manufacturing protocols and robust quality control systems are therefore needed to ensure batch-to-batch reproducibility and compliance with medical standards [98]. To address these challenges, adherence to international standards is paramount. Organizations such as the International Organization for Standardization (ISO) and ASTM International have

established comprehensive guidelines that govern the biological evaluation, mechanical testing, and quality management systems for medical devices [100]. For example, standards within the ISO 10993 series ensure the biocompatibility of PCL by providing protocols for in vitro and in vivo testing, while specific ASTM standards, such as F2902, offer guidance for the assessment of absorbable polymeric implants [101]. implementing these standards. manufacturers can establish robust quality control protocols that ensure batch-to-batch consistency in material properties and performance, thereby facilitating the transition from lab-scale prototypes to reliable, mass-produced medical devices [102].

Biomedical Applications of Polycaprolactone (PCL) Composites

8.2. Long-term Biocompatibility

Although PCL is generally well-tolerated and approved by regulatory agencies, the inclusion of additives, fillers, or surface modifications can alter its degradation profile and elicit unexpected immune responses over extended periods [44]. Comprehensive in vivo studies, including chronic implantation models, are essential to evaluate the safety and functionality of composite materials over their entire lifespan within the body.

8.3. Regulatory Issues

These issues also present considerable barriers to clinical translation. The multi-component nature of PCL-based composites often involving biologics, or nanoparticles places them in complex regulatory categories that vary between regions. The lack of standardized testing protocols and inconsistent classification systems can delay product approval and market entry [100,101]. There is a growing need for harmonized guidelines and collaborative efforts between researchers, industry stakeholders, and regulatory bodies to streamline the path from bench to bedside [102].

8.4. Integration with Digital Manufacturing

In the realm of manufacturing, the integration of PCL composites with digital fabrication technologies such as 3D and 4D printing represents a promising yet underexplored frontier [98,103]. These technologies allow for the creation of patient-specific implants and dynamic scaffolds that can change shape or function in response to environmental cues [103]. However, the development of PCL formulations that are both printable and retain desirable mechanical biological properties is still in its infancy. Further research is needed to optimize composite inks, printing parameters, and post-processing methods to fully leverage these innovations.

8.5. Multidisciplinary collaborations for next-gen **Applications**

Finally, the advancement of next-generation biomedical applications will require strong multidisciplinary collaboration. The convergence of materials science, biology, engineering, data science, and clinical medicine is essential for the design of intelligent, biofunctional PCL-based systems [103]. For instance, integrating machine learning algorithms for scaffold design or combining PCL composites with gene-editing tools like CRISPR demands coordinated expertise across domains. Funding mechanisms, shared research platforms, and translational consortia will play a critical role in fostering these collaborations and accelerating innovation in the field.

8.6. Intellectual Property Landscape

In the medical device industry, the intellectual property (IP) landscape is critical for commercial success, and PCL-based technologies are no exception [102]. Patents are a fundamental tool, providing companies with exclusive rights to their innovations, which in turn secures their market position and attracts crucial investment for research and development. This is especially true for complex technologies like PCL composites and 3D-printed implants, where a strong IP portfolio is a valuable assets [44]. The IP landscape for PCL includes patents on new materials, specific device designs, and innovative manufacturing processes, all of which are essential for navigating regulatory hurdles and achieving global market entry [103].

9. CONCLUSION

9.1. Summary of Key Findings

This review has comprehensively explored the properties, fabrication strategies, and biomedical Polycaprolactone (PCL)-based applications of composites. PCL's unique combination biocompatibility, biodegradability, mechanical flexibility, and FDA approval makes it a highly attractive base material for a wide range of biomedical uses. The incorporation of natural polymers, ceramics, and nanomaterials into PCL has enabled the development of multifunctional composites with enhanced biological performance, tailored degradation rates, and improved mechanical integrity. **Applications** in engineering, drug delivery, wound healing, and dental and orthopedic devices demonstrate the versatility and impact of PCL composites across diverse medical domains. Furthermore, the emergence of smart, gene-delivery stimuli-responsive systems and scaffolds highlights the adaptability of PCL platforms to meet next-generation biomedical demands.

9.2. Importance of PCL Composites in Advancing Biomedical Technology

PCL-based composites vital represent advancement in the design of bioengineered systems due to their ability to bridge material science and medical functionality. Their tunable properties allow for the creation of personalized, patient-specific devices, while their compatibility with advanced fabrication methods such as electrospinning and 3D printing positions them at the forefront of regenerative Additionally, the ease of chemical modification and capacity to host therapeutic agents PCL composites ideal candidates multifunctional biomedical platforms. These features collectively underscore the strategic role of PCL composites in driving innovation in tissue regeneration, targeted therapy, and bioresorbable medical devices.

9.3. Outlook on Future Development Paths

the future of PCL-based Looking ahead, composites lies in addressing current challenges while harnessing emerging technological frontiers. Focused efforts are needed improve long-term biocompatibility, standardize large-scale manufacturing, and navigate complex regulatory frameworks. The integration of digital manufacturing techniques such as 4D printing and the incorporation of biosensors, gene-editing tools, and smart drug release systems offer promising avenues for nextgeneration applications. Moreover, fostering collaboration multidisciplinary across materials science, bioengineering, data science, and clinical practice will be essential for translating laboratory innovations into real-world clinical solutions. With continued research and strategic development, PCL composites are poised to play a transformative role in the evolution of biomedical technologies.

FUNDING DECLARATION

This research received no specific grant from any funding agency in the public, commercial, or not-for-profit sectors.

COMPETING INTEREST DECLARATION

The authors declare that they have no competing interests.

AUTHOR CONTRIBUTIONS

Prajakta Subhedar: Conceptualization, Investigation, Writing-Original Draft

Dr. Divya Padmanabhan: Supervision, Final Approval

Dr. Richa Agrawal: Writing – Review & Editing, Visualization

REFERENCES

- [1] Anderson JM, Shive MS. Biodegradation and biocompatibility of PLA and PLGAmicrospheres. Adv Drug Deliv Rev 2012; 64: 72-82. https://doi.org/10.1016/j.addr.2012.09.004
- [2] Williams DF. On the mechanisms of biocompatibility. Biomaterials 2008; 29(20): 2941-53. https://doi.org/10.1016/j.biomaterials.2008.04.023
- [3] Place ES, George JH, Williams CK, Stevens MM. Synthetic polymer scaffolds for tissue engineering. Chem Soc Rev 2009; 38(4): 1139-51. https://doi.org/10.1039/b811392k
- [4] Woodruff MA, Hutmacher DW. The return of a forgotten polymer-Polycaprolactone in the 21st century. Prog Polym Sci 2010; 35(10): 1217-56. https://doi.org/10.1016/j.progpolymsci.2010.04.002
- [5] Cipitria A, Skelton A, Dargaville TR, Dalton PD, Hutmacher DW. Design, fabrication and characterization of PCL electrospun scaffolds-a review. J Mater Chem 2011; 21(26): 9419-53. https://doi.org/10.1039/c0jm04502k
- [6] Dash TK, Konkimalla VB. Poly-ε-caprolactone based formulations for drug delivery and tissue engineering: A review. J Control Release 2012; 158(1): 15-33. https://doi.org/10.1016/j.jconrel.2011.09.064
- [7] Ramalingam M, Young MF, Thomas V, Sun L, Chow LC, Nguyen K, et al. Polycaprolactone nanofibrous scaffolds for bone tissue engineering. Acta Biomater 2013; 9(5): 4859-72.
- [8] Mirkhani S, Kharaziha M, Karimzadeh F, Ramakrishna S. Electrospinning parameters and their effect on PCL nanofiber membranes for biomedical use. Mater Des 2023; 225: 111498.
- [9] Hettiaratchi MH, Giraud MO, Premaratne GS, McDevitt TC, Guldberg RE. Antioxidant PCL-based scaffolds for minimal transplant rejection. Biomaterials 2024; 304: 122165.
- [10] Dechy-Cabaret O, Martin-Vaca B, Bourissou D. Controlled ring-opening polymerization of lactones and related cyclic esters. Chem Rev 2004; 104(12): 6147-76. https://doi.org/10.1021/cr040002s
- [11] Pitt CG, Jeffcoat AR, Zweidinger RA, Schindler A. Sustained drug delivery systems. I. The characterization of poly(εcaprolactone). J Biomed Mater Res 1979; 13(3): 497-507. https://doi.org/10.1002/jbm.820130313
- [12] Lam CXF, Savalani MM, Teoh SH, Hutmacher DW. Dynamics of in vitro polymer degradation of polycaprolactone-based scaffolds: accelerated versus simulated physiological conditions. Biomed Mater 2008; 3(3): 034108. https://doi.org/10.1088/1748-6041/3/3/034108
- [13] Pitt CG, Chasalow FI, Hibionada YM, Klimas DM, Schindler Aliphatic polyesters. I. The degradation of poly(εcaprolactone) in vivo. J Appl Polym Sci 1981; 26(11): 3779-87
- [14] Sun H, Mei L, Song C, Cui X, Wang P. The in vivo degradation, absorption and excretion of PCL-based implant. Biomaterials 2006; 27(9): 1735-40. https://doi.org/10.1016/j.biomaterials.2005.09.019
- [15] Woodruff MA, Rath SN, Dalton PD, Hutmacher DW. The use of electrospun polycaprolactone scaffolds in soft tissue engineering and wound healing applications. Adv Drug Deliv Rev 2013; 65(4): 463-80.
- [16] Sivalingam G, Chattopadhyay S. Thermal degradation of poly(ε-caprolactone) and poly(ε-caprolactone-co-lactic acid). PolymDegrad Stab 2000; 69(1): 11-6. https://doi.org/10.1016/S0141-3910(02)00376-2
- [17] Schindler A, Pitt CG. Biodegradable polymers for sustained drug delivery. In: Takeru H, Robinson JR, editors. Sustained and Controlled Release Drug Delivery Systems. New York: CRC Press; 1978. p. 105-37.

- Sorrentino A, Gorrasi G, Vittoria V. Potential perspectives of [18] bio-nanocomposites for food packaging applications. Trends Food Sci Technol 2007; 18(2): 84-95. https://doi.org/10.1016/j.tifs.2006.09.004
- Zhao Y, Zhang Z, Pan Z, Liu Y. Surface modification of [19] polycaprolactone for tissue engineering: strategies and applications. Colloids Surf B Biointerfaces 2014; 123: 658-
- [20] Fu X, Xu M, Liu J, Qi C, Yan J, Sun H. Surface modification of polycaprolactone by plasma treatment and immobilization of biomolecules for biomedical applications. Polymers (Basel) 2020; 12(1): 93.
- Bacakova L, Filova E, Parizek M, Ruml T, Svorcik V. [21] Modulation of cell adhesion, proliferation and differentiation on materials designed for body implants. Biotechnol Adv 2011; 29(6): 739-67. https://doi.org/10.1016/j.biotechadv.2011.06.004
- Piskin E. Biodegradable polymers as biomaterials. J [22] Biomater Sci Polym Ed 1995; 6(9): 775-95. https://doi.org/10.1163/156856295X00175
- Middleton JC, Tipton AJ. Synthetic biodegradable polymers [23] as orthopedic devices. Biomaterials 2000; 21(23): 2335-46. https://doi.org/10.1016/S0142-9612(00)00101-0
- Athanasiou KA, Niederauer GG, Agrawal CM. Sterilization, [24] toxicity, biocompatibility and clinical applications of polylactic acid/polyglycolic acid copolymers. Biomaterials 1996; 17(2): 93-102. https://doi.org/10.1016/0142-9612(96)85754-1
- [25] Gentile P, Chiono V, Carmagnola I, Hatton PV. An overview of poly(lactic-co-glycolic) acid (PLGA)-based biomaterials for bone tissue engineering. Int J Mol Sci 2014; 15(3): 3640-59. https://doi.org/10.3390/ijms15033640
- [26] Makadia HK, Siegel SJ. Poly lactic-co-glycolic acid (PLGA) as biodegradable controlled drug delivery carrier. Polymers (Basel) 2011; 3(3): 1377-97. https://doi.org/10.3390/polym3031377
- Narayanan N, Jayaraman K, Sreenivasan K. Biodegradable [27] polymers: a review on synthesis, properties applications. Prog Biomater 2022; 11: 65-82.
- Saini P. Arora M. Kumar MNVR. Poly(lactic acid) blends in [28] biomedical applications. Adv Drug Deliv Rev 2016; 107: 47https://doi.org/10.1016/j.addr.2016.06.014
- Gomes ME, Ribeiro AS, Malafaya PB, Reis RL, Cunha AM. [29] A new approach based on injection moulding to produce biodegradable starch-based polymeric scaffolds: morphology, mechanical and degradation behaviour. Biomaterials 2001; 22(9): 883-9. https://doi.org/10.1016/S0142-9612(00)00211-8
- [30] Li WJ, Laurencin CT, Caterson EJ, Tuan RS, Ko FK. Electrospun nanofibrous structure: a novel scaffold for tissue engineering. J Biomed Mater Res 2002; 60(4): 613-21. https://doi.org/10.1002/jbm.10167
- Shim JH, Moon SH, Yun MJ, Jeon YC, Jeong CM, Cho DW, [31] et al. Stimulation of bone regeneration by bioactive hybrid scaffolds of PCL/HA and BMP-2-loaded microspheres. Biomaterials 2012; 33(33): 8579-90.
- Pitt CG, Gratzl MM, Jeffcoat AR, Zweidinger R, Schindler A. [32] Sustained drug delivery systems II: Factors affecting release rates from poly(ε-caprolactone) and related biodegradable polyesters. J Pharm Sci 1979; 68(12): 1534-8. https://doi.org/10.1002/jps.2600681219
- Sun H, Mei L, Song C, Cui X, Wang P. The in vivo [33] degradation, absorption and excretion of PCL-based implant. Biomaterials 2006; 27(9): 1735-40. https://doi.org/10.1016/j.biomaterials.2005.09.019
- U.S. Food and Drug Administration (FDA). Premarket [34] Approval (PMA) Database: Absorbable Sutures and Drug Delivery Devices. Silver Spring, MD: FDA; 2020. Available https://www.fda.gov/medical-devices/premarketapproval-pma-database
- Narayanan G, Vernekar VN, Kuyinu EL, Laurencin CT. [35] Polycaprolactone nanofibers for musculoskeletal tissue engineering. Acta Biomater 2016; 42: 133-46.

- [36] Mobini S, Specht CS, Hoque ME. Current trends in the clinical applications of polycaprolactone-based scaffolds in tissue engineering and regenerative medicine. Front BioengBiotechnol 2019; 7: 82.
- U.S. Food and Drug Administration (FDA). Material Safety [37] Data Sheets and Biocompatibility Summaries: Polycaprolactone. Silver Spring, MD: FDA; 2021. Available from: https://www.fda.gov/medical-devices/biocompatibilitymaterials
- Pitt CG, Chasalow FI, Hibionada YM, Klimas DM, Schindler [38] Aliphatic polyesters. I. The degradation of poly(εcaprolactone) in vivo. J Appl Polym Sci 1981; 26(11): 3779https://doi.org/10.1002/app.1981.070261124
- [39] Lam CXF, Savalani MM, Teoh SH, Hutmacher DW. Dynamics of in vitro polymer degradation of polycaprolactone-based scaffolds. Biomed Mater 2008; 3(3): 034108. https://doi.org/10.1088/1748-6041/3/3/034108
- Zhao Y. Zhang Z. Pan Z. Liu Y. Surface modification of [40] polycaprolactone for tissue engineering: strategies and applications. Colloids Surf B Biointerfaces 2014; 123: 658-67.
- [41] Fu X, Xu M, Liu J, Qi C, Yan J, Sun H. Surface modification of polycaprolactone by plasma treatment and immobilization of biomolecules for biomedical applications. Polymers (Basel) 2020; 12(1): 93.
- Sun H, Mei L, Song C, Cui X, Wang P. The in vivo degradation, absorption and excretion of PCL-based [42] implant. Biomaterials 2006; 27(9): 1735-40. https://doi.org/10.1016/j.biomaterials.2005.09.019
- Gentile P, Chiono V, Carmagnola I, Hatton PV. An overview [43] of poly(lactic-co-glycolic) acid (PLGA)-based biomaterials for bone tissue engineering. Int J Mol Sci 2014; 15(3): 3640-59. https://doi.org/10.3390/ijms15033640
- Rezwan K, Chen QZ, Blaker JJ, Boccaccini AR. Biodegradable and bioactive porous polymer/inorganic [44] composite scaffolds for bone tissue engineering. Biomaterials 2006; 27(18): 3413-31. https://doi.org/10.1016/j.biomaterials.2006.01.039
- [45] Li X, Xie J, Lipner J, Yuan X, Thomopoulos S, Xia Y. Nanofiber scaffolds with tunable structures, mechanical properties and biological functions. J Biomed Nanotechnol 2009; 5(4): 525-37.
- [46] Dash TK, Konkimalla VB. Poly-ε-caprolactone based formulations for drug delivery and tissue engineering: A review. J Control Release 2012; 158(1): 15-33. https://doi.org/10.1016/j.jconrel.2011.09.064
- Goonoo N, Bhaw-Luximon A, Passanha P, Esteves S, [47] Jhurry D. Polydioxanone-based bio-materials for tissue engineering and drug/gene delivery applications. Eur J Pharm Biopharm 2015; 97(Pt B): 371-91. https://doi.org/10.1016/j.ejpb.2015.05.024
- [48] Pitt CG, Jeffcoat AR, Zweidinger RA, Schindler A. Sustained drug delivery systems. I. The characterization of poly(εcaprolactone). J Biomed Mater Res 1979; 13(3): 497-507. https://doi.org/10.1002/jbm.820130313
- [49] Narayanan G, Vernekar VN, Kuyinu EL, Laurencin CT. Polycaprolactone nanofibers for musculoskeletal tissue engineering. Acta Biomater 2016; 42: 133-46.
- [50] Pego AP, Poot AA, Grijpma DW, Feijen J. Copolymers of trimethylene carbonate and ε-caprolactone for porous nerve guides: synthesis and properties. J Biomater Sci Polym Ed 2001; 12(1): 35-53. https://doi.org/10.1163/156856201744434
- [51] Chen JP, Chiang YJ. Bioactive electrospun silver nanoparticles-containing polyurethane nanofibers as wound dressings. J Nanosci Nanotechnol 2010; 10(11): 7560-4. https://doi.org/10.1166/jnn.2010.2829
- Bacakova L, Filova E, Parizek M, Ruml T, Svorcik V. [52] Modulation of cell adhesion, proliferation and differentiation on materials designed for body implants. Biotechnol Adv 2011; 29(6): 739-67. https://doi.org/10.1016/j.biotechadv.2011.06.004

- [53] Hutmacher DW, Schantz T, Zein I, Ng KW, Teoh SH, Tan KC. Mechanical properties and cell cultural response of polycaprolactone scaffolds designed and fabricated via fused deposition modeling. J Biomed Mater Res 2001; 55(2): 203-16. https://doi.org/10.1002/1097-4636(200105)55:2<203::AID-JBM1007>3.0.CO;2-7
- [54] Bose S, Roy M, Bandyopadhyay A. Recent advances in bone tissue engineering scaffolds. Trends Biotechnol 2012; 30(10): 546-54. https://doi.org/10.1016/j.tibtech.2012.07.005
- [55] Wang X, Li X, Ito Y, Sogo Y, Ohno K. Biodegradable polymer/β-tricalcium phosphate composites for tissue engineering scaffolds. Biomaterials 2003; 24(13): 2195-203.
- [56] Li JP, de Wijn JR, van Blitterswijk CA, de Groot K. Porous Ti6Al4V scaffold directly fabricating by rapid prototyping: preparation and in vitro experiment. Biomaterials 2006; 27(8): 1223-35. https://doi.org/10.1016/j.biomaterials.2005.08.033
- [57] Depan D, Girase B, Shah JS, Misra RDK. Structure– process–property relationship of the polar graphene oxidemediated cellular response and stimulated growth of osteoblasts on hybrid chitosan network structure nanocomposites. Acta Biomater 2011; 7(9): 3432-45. https://doi.org/10.1016/j.actbio.2011.05.019
- [58] Balani K, Anderson R, Laha T, Andara M, Tercero J, Crumpler E, et al. Plasma-sprayed carbon nanotube reinforced hydroxyapatite coatings and their interaction with human osteoblasts in vitro. Biomaterials 2007; 28(4): 618-24. https://doi.org/10.1016/j.biomaterials.2006.09.013
- [59] Saini P, Arora M, Kumar MNVR. Poly(lactic acid) blends in biomedical applications. Adv Drug Deliv Rev 2016; 107: 47-59. https://doi.org/10.1016/j.addr.2016.06.014
- [60] Makadia HK, Siegel SJ. Poly lactic-co-glycolic acid (PLGA) as biodegradable controlled drug delivery carrier. Polymers (Basel) 2011; 3(3): 1377-97. https://doi.org/10.3390/polym3031377
- [61] Athanasiou KA, Niederauer GG, Agrawal CM. Sterilization, toxicity, biocompatibility and clinical applications of polylactic acid/polyglycolic acid copolymers. Biomaterials 1996; 17(2): 93-102. https://doi.org/10.1016/0142-9612(96)85754-1
- [62] Ghorbani F, Moradi A, Shafiee A, Farokhi M. Recent advances in PLA-based biomaterials for tissue engineering applications. Mater Sci Eng C 2020; 110: 110958.
- [63] Geim AK, Novoselov KS. The rise of graphene. Nat Mater 2007; 6(3): 183-91. https://doi.org/10.1038/nmat1849
- [64] Goonoo N, Bhaw-Luximon A, Jhurry D. Polycaprolactone-based biomaterials for tissue engineering and drug/gene delivery applications: blending and functionalization. Int J Polym Mater Polym Biomater 2013; 62(2): 79-88.
- [65] Jones JR. Review of bioactive glass: from Hench to hybrids. Acta Biomater 2013; 9(1): 4457-86. https://doi.org/10.1016/j.actbio.2012.08.023
- [66] Rahaman MN, Day DE, Bal BS, Fu Q, Jung SB, Bonewald LF, et al. Bioactive glass in tissue engineering. Acta Biomater 2011; 7(6): 2355-73. https://doi.org/10.1016/i.actbio.2011.03.016
- [67] Li WJ, Laurencin CT, Caterson EJ, Tuan RS, Ko FK. Electrospun nanofibrous structure: a novel scaffold for tissue engineering. J Biomed Mater Res 2002; 60(4): 613-21. https://doi.org/10.1002/jbm.10167
- [68] Xue J, Wu T, Dai Y, Xia Y. Electrospinning and electrospun nanofibers: methods, materials, and applications. Chem Rev 2019; 119(8): 5298-415. https://doi.org/10.1021/acs.chemrev.8b00593
- [69] Murphy SV, Atala A. 3D bioprinting of tissues and organs. Nat Biotechnol 2014; 32(8): 773-85. https://doi.org/10.1038/nbt.2958
- [70] Bhardwaj N, Kundu SC. Electrospinning: a fascinating fiber fabrication technique. Biotechnol Adv 2010; 28(3): 325-47. https://doi.org/10.1016/j.biotechadv.2010.01.004

- [71] Li D, Xia Y. Electrospinning of nanofibers: reinventing the wheel? Adv Mater 2004; 16(14): 1151-70. https://doi.org/10.1002/adma.200400719
- [72] Sill TJ, von Recum HA. Electrospinning: applications in drug delivery and tissue engineering. Biomaterials 2008; 29(13): 1989-2006. https://doi.org/10.1016/j.biomaterials.2008.01.011
- [73] Labet M, Thielemans W. Synthesis of polycaprolactone: a review. Chem Soc Rev 2009; 38(12): 3484-504. https://doi.org/10.1039/b820162p
- [74] Goonoo N, Jeetah R, Bhaw-Luximon A, Jhurry D. Polycaprolactone-based biodegradable scaffolds for tissue engineering and drug delivery applications. J Biomed Mater Res A 2014; 102(10): 3649-70. https://doi.org/10.1166/jbn.2014.1885
- [75] Paul DR, Robeson LM. Polymer nanotechnology: nanocomposites. Polymer 2008; 49(15): 3187-204. https://doi.org/10.1016/j.polymer.2008.04.017
- [76] Shanmugam K, Kumar S, Shanmugam S. Polycaprolactone and its composites for biomedical applications. Mater Today Proc 2021; 46: 2706-11.
- [77] Saha S, Pal A, Paul S, Banerjee S. Polycaprolactone composites reinforced with strontium-substituted hydroxyapatite for bone regeneration. Mater Sci Eng C 2020; 108: 110420.
- [78] Zhang L, Gao L, Zhang Y, Zhang C, Xiao X, Li X, et al. ZnO-incorporated PCL/HA scaffolds for bone tissue regeneration. Mater Des 2021; 197: 109216.
- [79] Arcos D, Vallet-Regí M. Substituted hydroxyapatite coatings for bone tissue engineering. Acta Biomater 2020; 10(3): 927-36.
- [80] Jones JR. Review of bioactive glass: from Hench to hybrids. Acta Biomater 2013; 9(1): 4457-86. https://doi.org/10.1016/j.actbio.2012.08.023
- [81] Pego AP, Poot AA, Grijpma DW, Feijen J. Copolymers of trimethylene carbonate and ε-caprolactone for nerve guides: synthesis and properties. J Biomater Sci Polym Ed 2001; 12(1): 35-53. https://doi.org/10.1163/156856201744434
- [82] Wang Y, Shi X, Ren L. Biodegradable poly(trimethylene carbonate-co-caprolactone) copolymers and their biomedical applications. Polymers (Basel) 2018; 10(7): 739.
- [83] Melchels FPW, Feijen J, Grijpma DW. A review on stereolithography and its applications in biomedical engineering. Biomaterials 2010; 31(24): 6121-30. https://doi.org/10.1016/j.biomaterials.2010.04.050
- [84] Chimene D, Lennox KK, Kaunas RR, Gaharwar AK. Advanced bioinks for 3D printing: a materials science perspective. Ann Biomed Eng 2016; 44(6): 2090-102. https://doi.org/10.1007/s10439-016-1638-v
- [85] Bose S, Vahabzadeh S, Bandyopadhyay A. Bone tissue engineering using 3D printing. Mater Today 2013; 16(12): 496-504. https://doi.org/10.1016/j.mattod.2013.11.017
- [86] Williams JM, Adewunmi A, Schek RM, Flanagan CL, Krebsbach PH, Feinberg SE, et al. Bone tissue engineering using polycaprolactone scaffolds fabricated via selective laser sintering. Biomaterials 2005; 26(23): 4817-27. https://doi.org/10.1016/j.biomaterials.2004.11.057
- [87] Hutmacher DW, Schantz JT, Zein I, Ng KW, Teoh SH, Tan KC. Mechanical properties and cell cultural response of polycaprolactone scaffolds designed and fabricated via fused deposition modeling. J Biomed Mater Res 2001; 55(2): 203-16. https://doi.org/10.1002/1097-4636(200105)55:2<203::AID-</p>
 - https://doi.org/10.1002/1097-4636(200105)55:2<203::AID-JBM1007>3.0.CO;2-7
- [88] Chia HN, Wu BM. Recent advances in 3D printing of biomaterials. J Biol Eng 2015; 9: 4. https://doi.org/10.1186/s13036-015-0001-4
- [89] Loh QL, Choong C. Three-dimensional scaffolds for tissue engineering applications: role of porosity and pore size. Tissue Eng Part B Rev 2013; 19(6): 485-502. https://doi.org/10.1089/ten.teb.2012.0437

- [90] Derby B. Printing and prototyping of tissues and scaffolds. Science 2012; 338(6109): 921-6. https://doi.org/10.1126/science.1226340
- [91] Liu Y, Zheng Y, Li Y, Zhang X, Wang Y. Thermo- and pH-responsive PCL-based smart polymers for controlled drug delivery. Mater Sci Eng C 2021; 118: 111433.
- [92] Ranganathan S, Balagangadharan K, Selvamurugan N. Chitosan and PCL-based biomaterials in functionalized drug and gene delivery systems. Int J Biol Macromol 2019; 133: 354-65. https://doi.org/10.1016/j.ijbiomac.2019.04.115
- [93] Cao H, Xu S, Li X, Wang H, Zhang J, Xie M. Functionalized PCL nanofibrous scaffolds for targeted therapy and tissue engineering. Adv Funct Mater 2022; 32(12): 2108754.
- [94] Chen G, Wang Z, Wu J, Wang G, Xu T. PCL-based platforms for gene delivery: recent advances and future perspectives. J Control Release 2020; 322: 383-400.
- [95] Chia HN, Wu BM. Recent advances in 3D printing of biomaterials using machine learning for predictive scaffold design. J Biol Eng 2015; 9: 4. https://doi.org/10.1186/s13036-015-0001-4
- [96] Dutta S, Kadam S, Patel A, Roy S, Bandyopadhyay A. Machine learning-guided optimization of PCL composite scaffolds: predicting degradation and drug release. Mater Des 2022; 215: 110465.
- [97] Mobini S, Specht CS, Hoque ME. Clinical translation and market trends of polycaprolactone-based medical devices:

- from Neurolac® to Osteoplug®. Front BioengBiotechnol 2019; 7: 82.
- [98] Guvendiren M, Molde J, Soares RM, Kohn J. Designing biomaterials for 3D printing. ACS Biomater Sci Eng 2016; 2(10): 1679-93. https://doi.org/10.1021/acsbiomaterials.6b00121
- [99] He Y, Yang F, Zhao H, Gao Q, Xia B, Fu J. Research on the reproducibility of 3D printing of PCL scaffolds for biomedical applications. Biofabrication 2016; 8(2): 025009. https://doi.org/10.1088/1758-5090/8/3/035008
- [100] ISO 10993-1. Biological evaluation of medical devices- Part 1: Evaluation and testing within a risk management process. International Organization for Standardization; 2018.
- [101] ASTM F2902-16. Standard guide for assessment of absorbable poly (lactic-co-glycolic acid) and polycaprolactone for use in surgical implants. ASTM International; 2016.
- [102] Ratner BD, Hoffman AS, Schoen FJ, Lemons JE. Biomaterials Science: An Introduction to Materials in Medicine. 4th ed. San Diego: Academic Press; 2020. https://doi.org/10.1016/B978-0-12-816137-1.00001-5
- [103] Murphy SV, De Coppi P, Atala A. Opportunities and challenges of translational 3D bioprinting. Nat Biomed Eng 2020; 4(4): 370-80. https://doi.org/10.1038/s41551-019-0471-7

Received on 22-07-2025

Accepted on 21-08-2025

Published on 24-09-2025

https://doi.org/10.6000/1929-5995.2025.14.15

© 2025 Subhedar et al.

This is an open-access article licensed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the work is properly cited.